

PHYLOGENETIC EVIDENCE CONFIRMS *ADIANTUM MENGLIANENSE* AS A NEW RECORD FOR LAOS WITH THE REDISCOVERY OF *ADIANTUM GINGKOIDES* (PTERIDACEAE)

Phongphayboun Phonepaseuth^{1,2}

ABSTRACT

Adiantum menglianense is reported for Laos for the first time, based on a specimen collected in Luang Prabang Province, northern Laos, and supported by molecular evidence. Details on its habitat and distribution, and photographs, are provided. In addition, photographs of natural habitat and the phylogenetic position of *A. ginkgoides* C.Chr., an endemic and rare species from Laos, are provided for the first time as the species is rediscovered 95 years after its original collection.

Keywords: karst limestone, maidenhair fern, phylogeny, pteridophyte, *rbcL*

INTRODUCTION

Pteridophyte studies in Laos have recently gained attention. Between 2021 and 2025, approximately 36 species representing 27 genera and 12 families have been newly documented, including six species endemic to Laos, most of which are associated with limestone habitats (TAGANE *ET AL.* 2021; VONGTHAVONE *ET AL.*, 2021, 2025; SRIPENG *ET AL.*, 2024; ZHANG *ET AL.*, 2024; MIYAZAKI *ET AL.*, 2025). Limestone ecosystems in Laos are increasingly recognized as hotspots for endemic pteridophyte species as well as many newly documented species of flowering plants, e.g. *Begonia brunneosquamata* Phonep. & C.W.Lin (Begoniaceae) (PHONEPASEUTH & LIN, 2025), *Lysionotus calcicola* Phonep., Soulad. & Souvann. (Gesneraceae) (SOUVANNAKHOUUMANE *ET AL.*, 2025), *Tetrastigma fruticosum* Soulad., Phonep. & Tagane (TAGANE *ET AL.*, 2025), which are narrowly restricted to karst landscapes. As noted by LINDSAY & MIDDLETON (2012–, continuously updated), further exploration of limestone sites is likely to reveal additional locally endemic species, highlighting the importance of continued botanical surveys and conservation efforts in these habitats. Continued identification of herbarium collections and additional botanical surveys in poorly explored regions are likely to reveal further diversity within the fern flora of Laos. Details for each taxon of pteridophyte reported from Laos in past five years are provided in Appendix 1.

The genus *Adiantum* L., commonly known as “maidenhair ferns,” has a worldwide distribution and comprises approximately 245 accepted species (POWO, 2025). All species of *Adiantum* share a distinctive morphological feature not found in other fern lineages: their sporangia are borne directly on the reflexed margin of the pinnae, forming a “false

¹ Systematic Botany Laboratory, Department of Biological Sciences, Graduate School of Science, Tokyo Metropolitan University, 1-1 Minami-Osawa, Hachioji, Tokyo 192-0397, Japan.

² Faculty of Environmental Sciences, National University of Laos, Dongdok Campus, Xaythany District, Vientiane Capital, Laos. E-mail: p.phonepaseuth@nuol.edu.la

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indusium” or “pseudindusium” (TAGAWA & IWATSUKI, 1985; ZHANG *ET AL.*, 2013). Ten species of this genus have been recorded from Laos, namely *A. capillus-veneris* L., *A. caudatum* L., *A. erylliae* Tardieu & C.Chr., *A. flabellulatum* L., *A. ginkoides* C.Chr., *A. membranifolium* S. Linds. & Suksathan, *A. philippense* L., *A. soboliferum* Wall. ex Hook., *A. stenochlamys* Baker, and *A. zollingeri* Mett. ex Kuhn (LINDSAY & MIDDLETON, 2012–, continuously updated; SRIPENG *ET AL.*, 2024). Among them, *A. ginkoides* is the only species endemic to Laos, with little known about its habitat and distribution.

During field collections conducted in northern Laos (Luang Prabang and Ngoi districts, Luang Prabang Province) from August to September 2023, central Laos (Feuang District, Vientiane Province; Thakhek, Gnommalart, and Nakai districts, Khammouane Province) in August 2024; and northern and north-central Vietnam (Phu Tho and Nghe An provinces) in December 2024, several *Adiantum* species were collected, particularly those belonging to the morphologically challenging *A. philippense* complex. DNA barcoding is required to accurately distinguish species within this group.

EBIHARA *ET AL.* (2010) evaluated the utility of *rbcL* and *trnH-psbA* using 733 taxa of pteridophytes and found that these two barcodes are effective for identifying the pteridophyte flora of Japan. WANG *ET AL.* (2016) applied DNA barcodes to identify closely related species of Chinese *Adiantum* and found that *rbcL* had the highest PCR amplification rate (94.3%) and sequencing success rate (90.8%). Therefore, this study utilized *rbcL* as DNA barcoding to identify *Adiantum* species.

MATERIALS AND METHODS

DNA Extraction, Amplification and Sequencing

Total DNA was extracted from silica dried leaf material using a 2× CTAB protocol modified from DOYLE & DOYLE (1987). The *rbcL* gene (ribulose-1,5-bisphosphate carboxylase/oxygenase large subunit) was selected for barcoding in this study. This gene serves as a valuable phylogenetic marker for resolving deeper evolutionary relationships because of its relatively slow rate of evolution, and it is the most commonly sequenced plastid region in ferns (NITTA *ET AL.*, 2022).

For amplification of the chloroplast *rbcL* gene, I followed HORI *ET AL.* (2018), using primers namely af3 (5'-ATGTCACCACAAACGGAGACTAAAGC-3') and cr3 (5'-GCGGCAGCCAATTCGGACTCCA-3'). In addition, *trnL-F* intergenic spacer sequences including *trnL* intron (*trnL-L-F*) primers FernL1Ir1 (5'-GGYAATCCTGAGCCAAATC-3', LI *ET AL.* 2010) and F (5'-ATTTGAACTGGTGACACGAG-3', TABERLET *ET AL.*, 1991) were also amplified for *A. ginkoides* (accession number PX900790).

Polymerase chain reaction (PCR) amplifications were performed in a 10 µL reaction mixture containing 3.3 µL of ddH₂O, 0.6 µL of each forward and reverse primer (10 µM), 5 µL of Prime STAR Max DNA Polymerase (Takara, Kyoto, Japan) and 0.5 µL of DNA template. The PCR profile followed YONEOKA *ET AL.* (2024): 95 °C for 7 min, followed by 35 cycles of 98 °C for 10 s, 58 °C for 15 s, and 72 °C for 8 s, and 72 °C for 7 min. Amplification products were visualized under UV light after electrophoretic separation on a 1.5% agarose TAE gel stained with SYBR Safe gel stain (Applied Biosystems). To remove superfluous dNTPs and primers, PCR products were subsequently purified using ExoSAP-IT (Applied Biosystems) according to the manufacturer's protocols. Sequencing PCRs used the same primers as for amplification, and reaction mixtures for sequencing were prepared using the SuperDye Cycle Sequencing Kit v3.1 (Edge BioSystems, California, USA). The protocol included an initial denaturation at 96 °C for 45 s, followed by 28 cycles of denaturation at 96 °C for 10 s,

primer annealing at 50 °C for 5 s, and primer extension at 60 °C for 2 min. Sequencing PCR products were purified and analyzed at the Makino Herbarium using an AB 3130 Genetic Analyzer (Applied Biosystems).

Taxon Sampling and Phylogenetic Analyses

A total of 83 sequences were included in this study, 82 of *Adiantum* spp. as ingroup taxa, and *Antrophyum latifolium* Blume (vittarioids) as the outgroup taxon, following HUIET *ET AL.* (2018). Besides that, 8 sequences of 6 species were newly generated from this study (*A. ginkgoides* is sequenced for the first time) (See Table 1). The remaining sequences were downloaded from GenBank (see Table 2), including most species in the *A. philippense* clade and species that occur in Eurasia (HUIET *ET AL.*, 2018) with additional samples from previous literature (EBIHARA *ET AL.*, 2010; LU *ET AL.*, 2012; LESTARI *ET AL.*, 2014; KUO *ET AL.*, 2016; WANG *ET AL.*, 2016). The sequence alignment was performed with MAFFT (KATO *ET AL.*, 2019) followed by manually checking and editing in Aliview (LARSSON, 2014). A maximum likelihood tree was constructed and analyzed using the IQ-TREE web server (version 1.6.12) (NGUYEN *ET AL.*, 2015), with TIME+I+G4 for *rbcL* model selected according to Bayesian Information Criterion (BIC), ModelFinder (KALYANAMOORTHY *ET AL.*, 2017) and employing one-thousand bootstrap pseudoreplicates via the ultrafast bootstrap (HOANG *ET AL.*, 2018). The tree was visualized using FigTree v.1.4 (RAMBAUT, 2012).

Table 1. Taxon, country of origin, collection location and voucher specimens of newly sequenced in this study

Taxon	Country	Locality	Voucher (herbarium code)	GenBank accession number (<i>rbcL</i>)
<i>Adiantum ginkgoides</i> C.Chr.	Laos	Gnommalart District, Khammouane Province	Phonepaseuth, P. et al. P097 (FOF)	PX900791
<i>Adiantum</i> sp. LPB23025	Laos	Ngoi District, Luangprabang Province	Phonepaseuth, P. & Vongsavath, V. LPB23025 (FOF)	PX900794
<i>Adiantum</i> sp. LPB23018	Laos	Ngoi District, Luangprabang Province	Phonepaseuth, P. & Vongsavath, V. LPB23018 (FOF)	PX900798
<i>Adiantum</i> sp. FE23002	Laos	Feuang District, Vientiane Province	Phonepaseuth, P. et al. FE23002 (FOF)	PX900793
<i>Adiantum</i> sp. FE23005	Laos	Feuang District, Vientiane Province	Phonepaseuth, P. et al. FE23005 (FOF)	PX900792
<i>Adiantum</i> sp. NK028	Laos	Nakai District, Khammouane Province	Phonepaseuth, P. et al. NK028 (FOF)	PX900795
<i>Adiantum</i> sp. PHVN012	Vietnam	Pu Hoat Nature Reserve, Nghe An Province.	Phonepaseuth, P. et al. PHVN012 (IEBR)	PX900796
<i>Adiantum</i> sp. PHVN059	Vietnam	Pu Hoat Nature Reserve, Nghe An Province.	Phonepaseuth, P. et al. PHVN059 (IEBR)	PX900797

Table 2. Ingroup and outgroup taxa sampled in this study. For each collection, the species name, Country of origin, voucher information, and GenBank accession numbers (*rbcL*) are provided.

Taxon	Country	Voucher (herbarium code)	GenBank number
<i>A. calcareum</i> Gardner	Brazil	Hirai 717 (SP)	MH019545
<i>A. capillus-junonis</i> Rupr.	China	Sundue 988 (DUKE)	MH019547
<i>A. capillus-junonis</i> Rupr.	China (Guangxi)	J.-M. Lu 111 (KUN)	JF935314
<i>A. capillus-junonis</i> Rupr.	Japan (Oita Pref.)	743693 (TNS)	AB574795
<i>A. capillus-veneris</i> L.	U.S.A.	Huiet 104 (UC)	KU147273
<i>A. capillus-veneris</i> L.	Russia	J. Wen 10360 (US)	JF935345
<i>A. capillus-veneris</i> L.	Japan (Oita Pref.)	777847 (TNS)	AB574796
<i>A. capillus-veneris</i> L.	China (Guangxi)	J.-M. Lu 138 (KUN)	JF935320
<i>A. caudatum</i> L.	Laos	Sugong WS-1837 (MO)	MH019548
<i>A. caudatum</i> L.	China (Hainan)	J.-M. Lu 209 (KUN)	JF935296
<i>A. caudatum</i> L.	China (Yunnan)	J.-M. Lu 050 (KUN)	JF935297
<i>A. caudatum</i> L.	India	Shyamalima Saikia	PP801084
<i>A. comorense</i> Fée	Tanzania	Mwangoka 5695 (MO)	MH019551
<i>A. davidii</i> Franch.	China	Kuo 1561 (TAIF)	MH019554
<i>A. deflectens</i> Mart.	Brazil	Schuettpelz 1390 (DUKE)	MH019557
<i>A. delicatulum</i> Mart	Brazil	Hirai 704 (SP)	MH019558
<i>A. diaphanum</i> Blume	Taiwan	777847 (TNS)	AB574797
<i>A. diaphanum</i> Blume	China	Wuzhishan Fern Survey 67 (MO)	MH019560
<i>A. diaphanum</i> Blume	China (Hainan)	J.-M. Lu 208 (KUN)	JF935301
<i>A. edgeworthii</i> Hook.	Japan (Oita Pref.)	743692 (TNS)	AB574798
<i>A. edgeworthii</i> Hook.	Vietnam	Kuo 1840 (TAIF)	MH019565
<i>A. flabellulatum</i> L.	Japan (Okinawa Pref.)	764008 (TNS)	AB574799
<i>A. flabellulatum</i> L.	Taiwan	Schuettpelz 1016A (DUKE)	MH0147275
<i>A. flabellulatum</i> L.	Taiwan	Kuo 3377 (TAIF)	PP559815
<i>A. flabellulatum</i> L.	China (Guizhou)	J.-M. Lu 192 (KUN)	JF935315
<i>A. flabellulatum</i> L.	China (Guangdong)	J.-M. Lu 450 (KUN)	JF935295
<i>A. flabellulatum</i> L.	China (Hainan)	J. Wen 6585 (US)	JF935325
<i>A. formosanum</i> Tagawa	Taiwan	Li 1402 (DUKE)	MH019571
<i>A. formosum</i> R.Br.	New Zealand	J. Wen 10779 (US)	JF935352
<i>A. formosum</i> R.Br.	-	Cultivated, Smith s.n. (UC)	KC984520
<i>A. gravesii</i> Hance	China	Zhang 376 (MO)	MH019579
<i>A. gravesii</i> Hance	China (Guangxi)	J.-M. Lu 129 (KUN)	JF935312
<i>A. gravesii</i> Hance	China (Guangdong)	J.-M. Lu 451 (KUN)	JF935317
<i>A. hispidulum</i> Sw.	Taiwan	Schuettpelz 1168A (DUKE)	MH019580
<i>A. hosei</i> Baker	Malaysia	Beaman 10302 (UC)	MH019582
<i>A. induratum</i> H.Christ.	Vietnam	WP 963 (MO)	MH019585
<i>A. induratum</i> H.Christ.	China (Hainan)	J.-M. Lu 210 (KUN)	JF935309
<i>A. induratum</i> H.Christ.	-	isolate WFH008	KT426956
<i>A. incisum</i> Forssk.	Tanzania	Gereau 6840 (MO)	MH019584
<i>A. madagascariense</i> H.Rosend.	Madagascar	Trigui 531 (MO)	MH019594
<i>A. malesianum</i> J.Ghatak	Malaysia	Schuettpelz 666 (DUKE)	MH019596
<i>A. malesianum</i> J.Ghatak	China (Guangdong)	L.Huiet 111 (UC)	EF452132
<i>A. mariesii</i> Baker	China	Wade 2493 (TAIF)	MH019597
<i>A. mariesii</i> Baker	China (Guangxi)	J.-M. Lu 120 (KUN)	JF935302
<i>A. menglianense</i> Y.Y.Qian	Taiwan	MO Awan-1 (TAIF)	KU945417
<i>A. menglianense</i> Y.Y.Qian	-	Yan12415	KP637234
<i>A. menglianense</i> Y.Y.Qian	-	Yan12409	KP637233

Table 2 (continued).

Taxon	Country	Voucher (herbarium code)	GenBank number
<i>A. menglianense</i> Y.Y.Qian	China (Guangxi)	J.-M. Lu 114 (KUN)	JF935311
<i>A. mettenii</i> Kuhn	Tanzania	Massawe 364 (MO)	MH019602
<i>A. monochlamys</i> D.C.Eaton	Taiwan	Li 427 (TAIF)	MH019603
<i>A. monochlamys</i> D.C.Eaton	Japan (Tokyo Pref.)	764001 (TNS)	AB574800
<i>A. myriosorum</i> Baker	China	Kuo 2128 (TAIF)	MH019605
<i>A. ogasawarense</i> Tagawa	Japan (Tokyo Pref.)	774843 (TNS)	AB574801
<i>A. pedatum</i> L.	U.S.A.	Rothfels 3839 (DUKE)	MH019612
<i>A. pedatum</i> L.	-	Huiet 117 (UC DNA voucher)	U05602
<i>A. pedatum</i> L.	Japan (Shizuoka Pref.)	764338 (TNS)	AB574802
<i>A. phanerophlebium</i> (Bak.) C.Chr.	Madagascar	J. Kluge 8039 (Z)	MH019615
<i>A. philippense</i> L.	Costa Rica	Rothfels 2655 (DUKE)	MH019616
<i>A. philippense</i> L.	Indonesia	WN153	LC004122
<i>A. philippense</i> L.	Indonesia	WN142	LC004111
<i>A. philippense</i> L.	Indonesia	J. Wen 10206 (US)	JF935340
<i>A. philippense</i> L.	Philippines (Luzon)	J. Wen 8257 (US)	JF935330
<i>A. philippense</i> L.	China (Yunnan)	J.-M. Lu 001 (KUN)	JF935331
<i>A. philippense</i> L.	China (Yunnan)	Y.-X. Zhang 009 (KUN)	JF935299
<i>A. poiretii</i> Wikstr.	Mexico	Windham 513 (DUKE)	MH019620
<i>A. schweinfurthii</i> Kuhn	Tanzania	Gereau 5966 (MO)	MH019630
<i>A. soboliferum</i> Wall. ex Hook.	Laos	S.K. Wu 294 (MO)	MH019635
<i>A. soboliferum</i> Wall. ex Hook.	-	isolate WFH038	KT427017
<i>A. soboliferum</i> Wall. ex Hook.	-	isolate Sunhang17076	KT427016
<i>A. stenochlamys</i> Baker	Malaysia	Schuettpelz 900A (DUKE)	MH019636
<i>A. taiwanianum</i> Tagawa	Taiwan	Schuettpelz 1131A (DUKE)	MH019637
<i>A. thongthamii</i> Suksathan	Thailand	Suksathan 3303 (QBG)	MH019639
<i>A. rhizophyllum</i> Schrad.	Brazil	Fiaschi 3568 (SPF)	MH019626
<i>Antrophyum latifolium</i> Blume	Papua New Guinea	Ranker 1774 (COLO)	EF452138

RESULTS

Phylogenetic Analyses

The aligned dataset contained 83 sequences and 1,309 nucleotide sites, of which 973 (74.3%) were constant and 278 (21.2%) were parsimony informative. The ML tree (Fig. 1) shows the phylogenetic relationships of eight newly collected specimens of *Adiantum* spp. Six specimens of *Adiantum* from Laos were resolved within the “philippense clade” and were distributed among several strong supported subclades (UFBS = 95–100). *Adiantum* sp. LPB23018 was placed in a strongly supported subclade together with accessions identified as *A. menglianense* from China and Taiwan (UFBS = 95), and was clearly separated from the clade containing *A. philippense* sensu stricto (UFBS = 94). Two Laotian specimens (*Adiantum* sp. NK028, *Adiantum* sp. FE23002) clustered with accessions of *A. philippense*, whereas *Adiantum* sp. LPB23025 and *Adiantum* sp. FE23005 were placed with *A. caudatum* and *A. soboliferum*, respectively, each with strong bootstrap support (UFBS = 98–100). *Adiantum ginkgoides* C.Chr. formed a strongly supported sister relationship with *A. capillus-junonis* Rupr. in the ML tree with strong bootstrap support (UFBS = 100) within

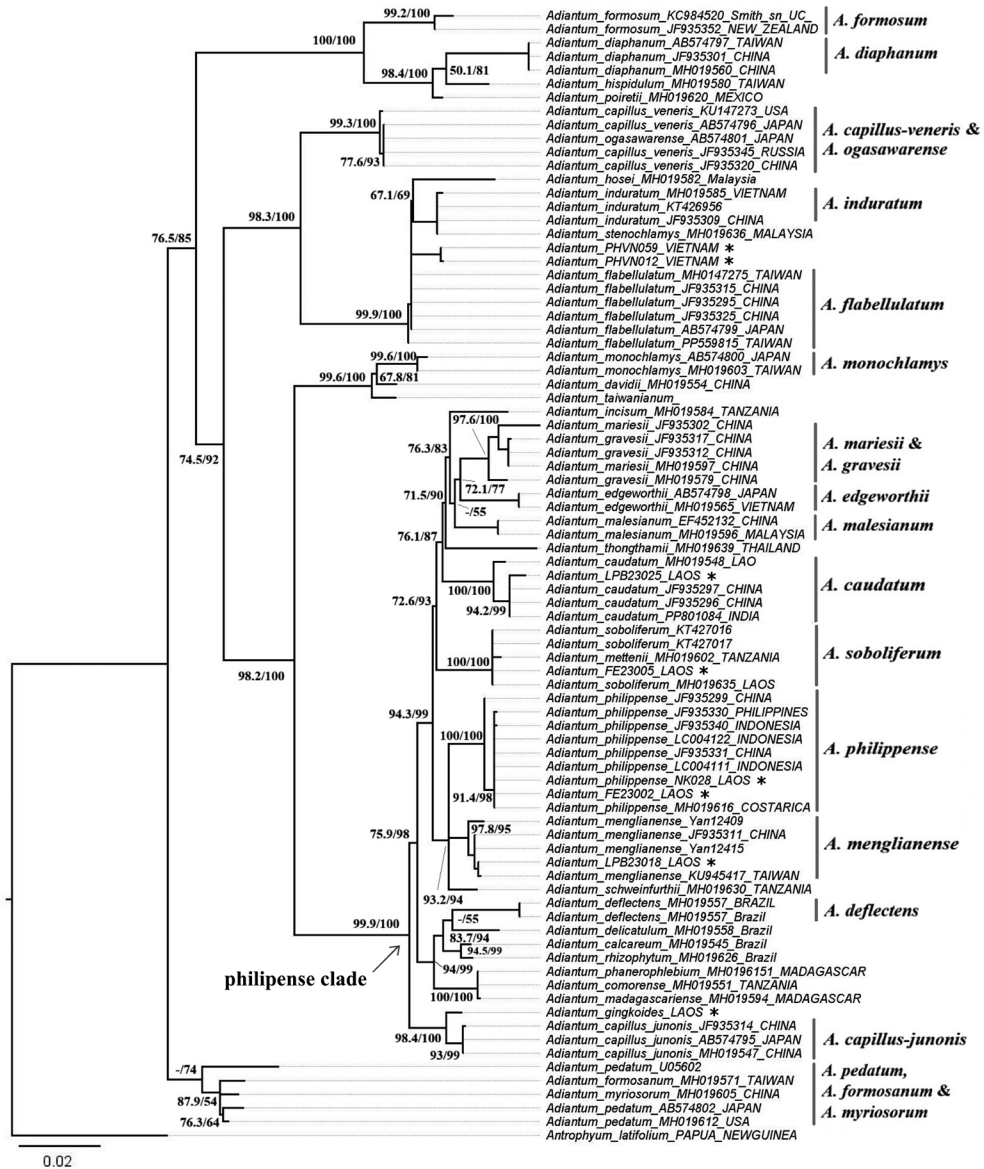


Figure 1. Maximum likelihood trees of species of *Adiantum* based on *rbcL* sequences. Species shown with asterisk are newly sequences in this study. Bootstrap values including Shimodaira-Hasegawa-like approximate likelihood ratio test (SH-aLRT) and Ultrafast bootstraps (UFBS) are shown at each branch (SH-aLRT/UFBS) when both values exceed 50%. Values greater than 80% for SH-aLRT and 95% for UFBS are considered as strong support.

the philipense clade and was distant from morphologically similar taxa such as *A. gravesii*. The two Vietnamese specimens of *Adiantum* cf. *flabellulatum* (PHVN059, PHVN012) were resolved within a broader clade including *A. flabellulatum*, and *A. hosei*, *A. induratum*, *A. stenochlamys* clade.

Taxonomic Account

Adiantum menglianense Y.Y.Qian [Pteridaceae] (Fig. 2)

Acta Bot. Austro Sin. 8: 37–38 (1992).

Type: CHINA. Yunnan: Menglian, 900–1,400 m, 23 December 1984, *Qian 801* (lectotype: HITBC-81381, designated by KUO *ET AL.* 2016; islectotypes: HITBC-81380, HITBC-109908).

This species is placed in the *Adiantum philippense* complex (KUO *ET AL.*, 2016). First described from Yunnan Province, South China (QIAN, 1992), and also confirmed to occur in Taiwan with phylogenetic analysis, determined by reproductive mode and ploidy level, in which 64 spores per sporangium and sexual diploids were found (KUO *ET AL.*, 2016). This species has more dissected fertile pinnules than *A. philippense* L. with narrower false indusia 2–6 mm (vs. *A. philippense* subsp. *philippense* (2–12 mm) (QIAN, 1992; KUO *ET AL.*, 2016).

Habitat and ecology: This species was found growing on red soil mixed with degraded limestone in a shaded semi-deciduous forest at an altitude of 440–460 m a.s.l. It shares its habitat with *A. caudatum* L. and other flowering plants such as *Begonia luangprabangensis* Phoney., and *B. aff. demissa* Craib (Begoniaceae), *Microchirita* sp. (Gesneriaceae).

Distribution: China (Yunnan Province), Laos (Luang Prabang Province), Taiwan.

Specimens examined: LAOS. Luang Prabang Province, Ngoi District, Nongkhiauw village, on the walking trail to Nangnone karst viewpoint. Around point 20°34'33"N, 102°36'30"E, 440–460 m a.s.l. 23 August 2023, *Phonepaseuth, P. & Vongsavath, V. LPB23018* (FOF). Same Province, Luang Prabang District, Long Lan mountain, along trail from Ban Den Savang to Ban Long Vai, 750 m a.s.l., 24 November 2017, *Hsu, T.C. 9982* (TAIF [TAIF540938, digital image!]).

Adiantum ginkgoides C.Chr. [Pteridaceae] (Fig. 3)

Bull. Mus. Natl. Hist. Nat., Sér. 2, 6: 100 (1934).

Type: LAOS. Thakhek district, Tham Village, on the limestone walls of a very humid cave, December 1930. *Colani, M. 4139* (holotype BM, isotype NY [NY00127470, digital image!], P [P00607853, P00607854, digital image!], UC [UC74445, digital image!], US [US00134480, digital image!]).

This species has fronds resembling those of *Ginkgo biloba* L. (Ginkgoaceae), but noted misspelling of epithet (LINDSAY & MIDDLETON, 2012–, continuously updated). This species differs from closely related species in having a darker and narrower indusium, non-proliferous leaves, shorter petioles, and margin of the pinnae not being incised between the sori (CHRISTENSEN, 1934). The type specimens were collected from “Village de Tham, Common Prov.” in December 1930 by the French archaeologist Madeleine Colani. Based on the map of villages and schools provided by the Ministry of Education and Sports of Laos (MOES, 2024), this locality is presumed to be the Elephant Cave, a notable large cave in present day Thakhek District, Khammouane Province, Laos. Additional evidence indicates that Colani was working in Thakhek district, Khammoune Province during October 1930 to studied vestiges of Bacsonian-Hoabinhian, for the Neolithic traces (GENOVESE, 2015: 60). In January 1931, Colani collected another set of *A. ginkgoides* specimens, together with *Antrophyum* species, in “Village de Mahaxay, Mahaxay District” (see specimens examined), which corresponded with her documented activities during that month (GENOVESE, 2023:

152). Since Colani was not a plant taxonomist, she may have passed these specimens to the French botanist and entomologist Paul Alfred Pételot, who then assigned herbarium numbers. Consequently, some of the type specimens collected by her are cited under Pételot's name rather than Colani's.

This species was rediscovered and photographed by me on 18 August 2019, in a field survey in Gnommalart District, Khammouane Province, at the time an immature plant (without spores, Fig. 3E). A subsequent collection from the same locality was made three years later (Fig. 3F–H).

Based on morphology notes provided by CHRISTENSEN (1934), this species is closely related to *A. leveillei* Christ (currently treated as *A. gravesii* Hance) and *A. semiorbiculatum* Bonap., but differs in having fewer and larger pinnae, shorter pinna petioles, and non-glaucous fronds. Examination of newly collected specimens, together with comparison to the type material, line drawings and published descriptions (CHRISTENSEN, 1934; TARDIEU-BLOT & CHRISTENSEN, 1938; LINDSAY & MIDDLETON, 2012–, continuously updated) confirm the overall match with *Adiantum ginkgoides*.

Habitat and ecology: This species grows on humid limestone walls and near cave entrances from altitudes of 150 m to 240 m a.s.l. It shares its habitat with *Argostemma* spp. (Rubiaceae), *Impatiens* sp. (Balsaminaceae), *Begonia murina* Craib (Begoniaceae), *Microchirita hamosa* (R.Br.) Yin Z.Wang and *Paraboea khotamiae* Phonep. & Souvann. (Gesneriaceae).

Distribution: Endemic to Laos, currently known from four localities in three districts (Thakhek, Gnommalart, and Mahaxay) of Khammouane Province.

Proposed conservation status: This species was found only in Khammouane Province, Central Laos. Numbers of individuals from 20 to 100 were found in new localities (Pha Kataiythong karst area, and Ban Mouang karst area, respectively). The Area of Occupancy (AOO) is calculated at 16 km² and the Extent of Occurrence (EOO) at 349.98 km² using GeoCat (BACHMAN *ET AL.*, 2011). Suitable habitat for this species extends into Phou Hin Poun National Park. However, several limestone karsts outside the national park are being destroyed for cement production and road construction. Based on these data, the species is preliminarily assessed as Endangered (EN, B1ab(iii); B2ab(iii); D).

Specimens examined: LAOS. Khammouane Province, Gnommalart District (new district record), Pha Kataithong, 17°33'09"N, 105°05'50"E, at altitude 225 m a.s.l., 16 September 2022, *Phonepaseuth, P. et al. P097* (FOF); Same district, Ban Muang karst limestones, 17°32'37"N, 105°03'55"E, at altitude 230–240 m a.s.l., 5 September 2024, *Tagane et al. Z1675* (FOF, KAG [KAG188338, digital image!]). Mahaxay District, Mahaxay village, on limestone rock in humid tall forest, at 160 m a.s.l., 5 January 1931, *Colani, M. 4252* (P [P01294135, digital image!], MICH [MICH1592761, digital image!]). *ibid.*, at 150 m a.s.l., 22 January 1931, *Colani, M. 4256* (P [P01462195, digital image!], MICH [MICH1592760, digital image!]).

DISCUSSION

Taxonomic Interpretation of Phylogenetic Results

Adiantum menglianense as a new record for Laos

The Laotian specimen LPB23018 was placed with strong support within the clade containing reference sequences of *Adiantum menglianense* from China and Taiwan, and was clearly separated from *A. philippense* *sensu stricto*. While this molecular result alone is

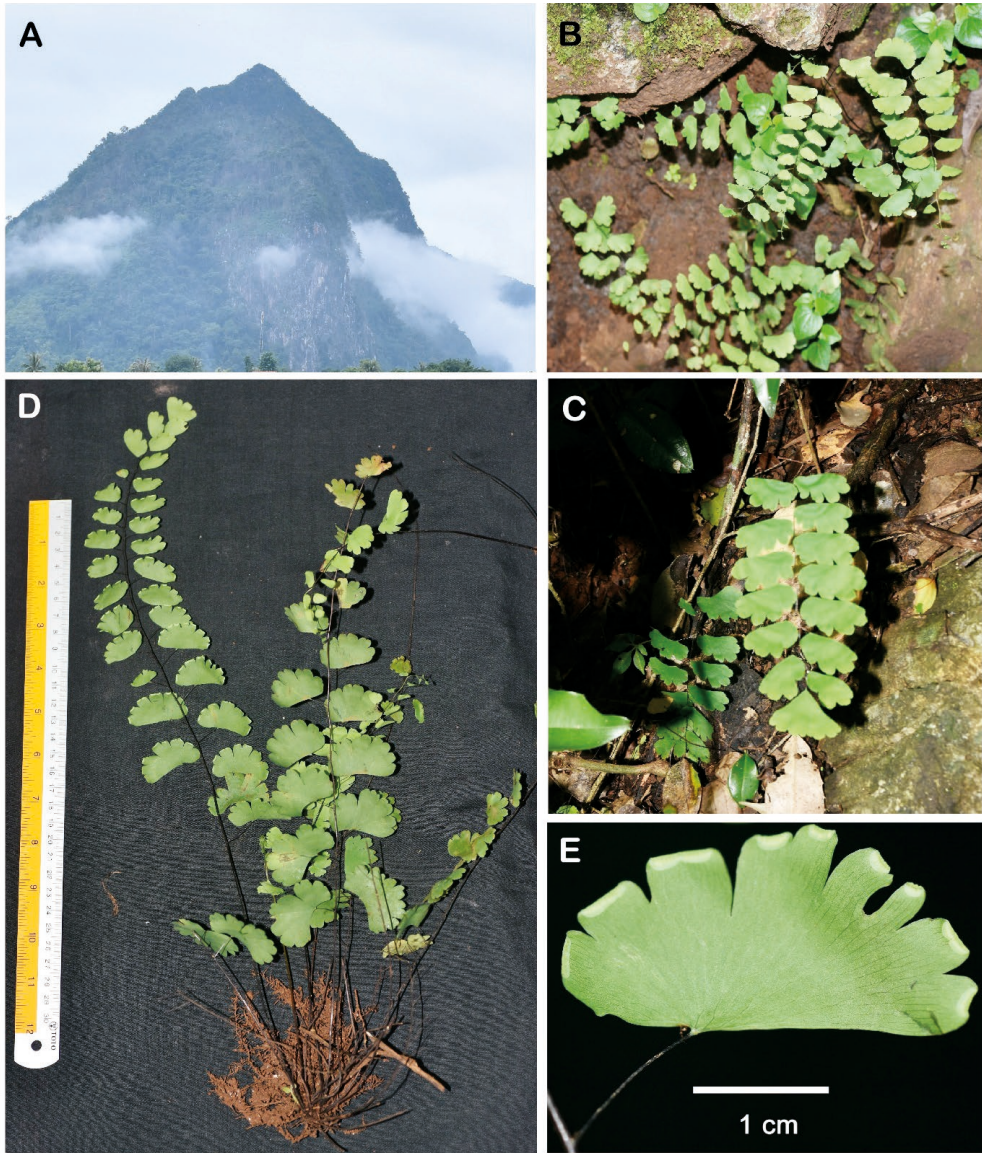


Figure 2. *Adiantum menglianense* Y.Y.Qian. A, habitat; B–C, plant habit in habitat; D, whole plant; E, portion of lower pinna surface showing false indusia. Photos by Phongphayboun Phonpaseuth (taken on 23 August 2023 at Nangnone karst viewpoint, Ngoi District, Luang Prabang Province).

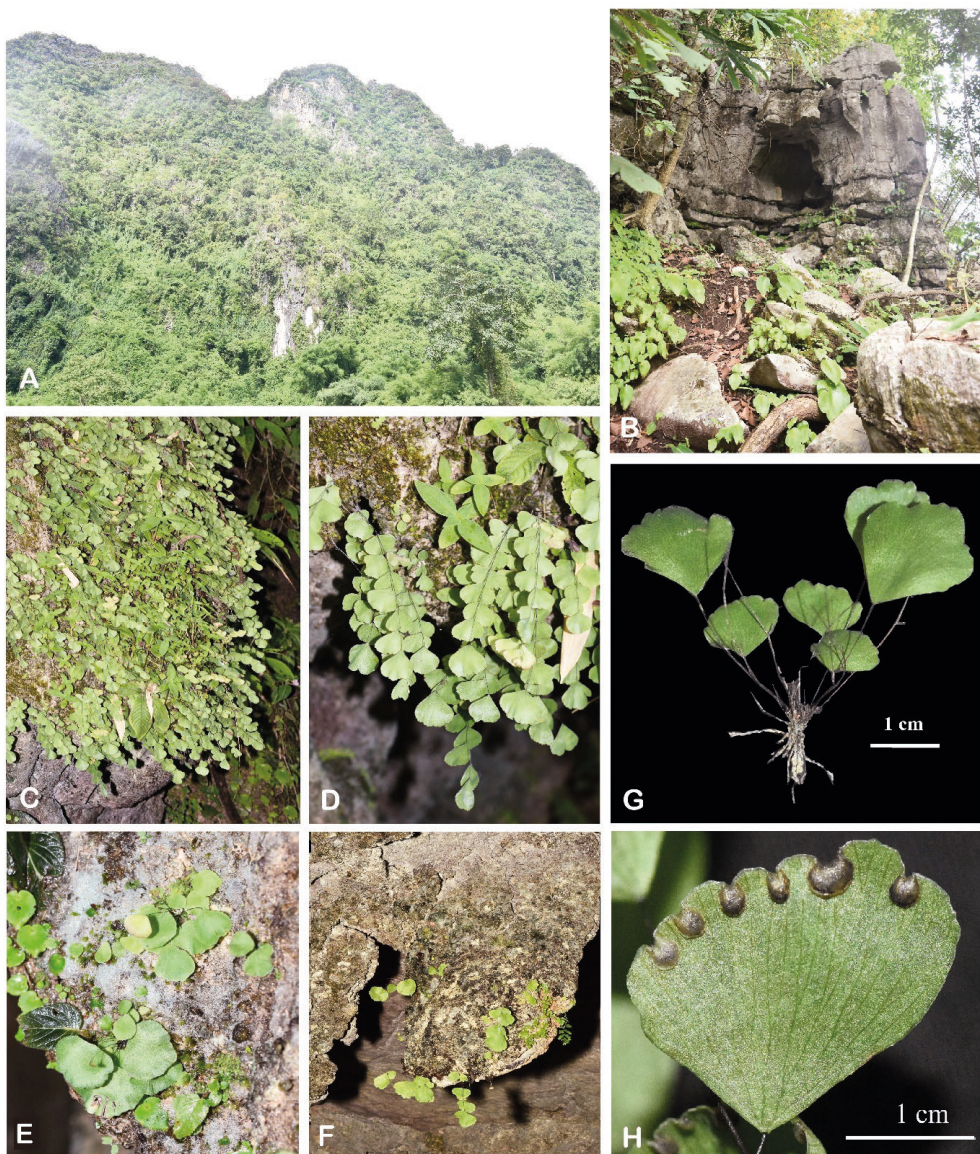


Figure 3. *Adiantum ginkgoides* C.Ch. A–B, habitat; C–F, habit in natural habitat (C, on wet limestone wall; D–F, on caves entrance); G, whole plant; H, portion of lower pinna surface showing false indusia. Photos by Phongphayboun Phonepaseuth (A–D taken on 5 September 2024 in Ban Muang karst limestones, Gnommalart District, Khammoun Province; E, taken on 18 August 2019; F–H, on 16 September 2022 in Pha Kataithong, Gnommalart District, Khammoun Province).

insufficient to confirm species identity, morphological examination of the Laotian material provides congruent evidence.

The specimens exhibit diagnostic characters consistent with *A. menglianense*, including more deeply dissected fertile pinnules (5–10 segments) and narrower false indusia (1.8–5.8 mm), which differ from those of *A. philippense* as documented by QIAN (1992) and KUO *ET AL.* (2016). The agreement between molecular placement and morphological characters supports the interpretation that the Laotian population represents *A. menglianense*. On this basis, the species is here recognized as newly recorded for the fern flora of Laos.

Rediscovery and phylogenetic position of Adiantum ginkgoides

Adiantum ginkgoides, a species endemic to Laos and not collected since its original discovery in 1930–1931, was rediscovered during recent field surveys in Khammouane Province. Morphological examination of newly collected specimens, together with comparison to type material, historical descriptions, and published illustrations (CHRISTENSEN, 1934; TARDIEU-BLOT & CHRISTENSEN, 1938; LINDSAY & MIDDLETON, 2012–, continuously updated), confirms their close correspondence with *A. ginkgoides*.

Phylogenetically, *A. ginkgoides* was resolved within the philippense clade and formed a strongly supported sister relationship with *A. capillus-junonis*. This placement contrasts with earlier assumptions of close affinity with morphologically similar species such as *A. gravesii*. The combined morphological and molecular evidence supports the continued recognition of *A. ginkgoides* as a distinct species and clarifies its evolutionary relationships within *Adiantum*.

In several cases, including specimens provisionally identified as *Adiantum* cf. *flabellulatum*, molecular data alone were insufficient to resolve species identity, indicating that detailed morphological examination remains necessary for confident identification.

DNA Barcoding and Species Identification in the Laotian Fern Flora

DNA barcoding has become an essential tool for resolving species boundaries in taxonomically challenging fern groups, particularly within *Adiantum* (LESTARI *ET AL.*, 2014; KUO *ET AL.*, 2016; WANG *ET AL.*, 2016; HUIET *ET AL.*, 2018). In this study, molecular evidence was essential for confirming the presence of *A. menglianense* in Laos for the first time, as well as for clarifying the phylogenetic position of the endemic *A. ginkgoides*, a species not observed since its original collection 95 years ago. These results highlight how genetic data provide reliable diagnostic characters where morphology alone is insufficient, and demonstrate the value of integrating DNA barcodes into taxonomic research on the fern flora of Laos.

Among 332 fern and lycophyte specimens collected from 2022 to 2024, 34 specimens (approximately 10%) were initially identified as unrecorded or unknown from Laos. These were mainly collected from limestone areas in northern and central Laos, as well as from regions with limited prior sampling in southern Laos, e.g. Dak Cheung Plateau (PHONEPSEUTH *ET AL.*, 2024). Species identification and DNA barcoding of these specimens are currently ongoing. This work represents an important first step toward establishing a genetic database of fern species in Laos to support future research and conservation efforts.

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REFERENCES

- BACHMAN, S., J. MOAT, A.W. HILL, J. DE LA TORRE, AND B. SCOTT. 2011. Supporting Red List threat assessments with GeoCAT: geospatial conservation assessment tool. *Zookeys* 150: 117–126. <https://doi.org/10.3897/zookeys.150.2109>
- CHRISTENSEN, C. 1934. Filices Novae Indochinenses. *Bull. Mus. Natl. Hist. Nat., Sér.* 2, 6: 100–106.
- DOYLE, J. J., AND J. L. DOYLE. 1987. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. *Phytochem. Bull.* 19: 11–15.
- EBIHARA, A., J. H. NITTA, AND M. ITO. 2010. Molecular species identification with rich floristic sampling: DNA barcoding the pteridophyte flora of Japan. *PLoS One*. 5 (12): e15136. <https://doi.org/10.1371/journal.pone.0015136>.
- GENOVESE, R. 2015. *The plain of jars of north Laos: beyond Madeleine Colani* [PhD thesis, SOAS, University of London]. <https://doi.org/10.25501/SOAS.00020397>
- GENOVESE, L. 2023. Katherine Routledge and Madeleine Colani: Victorian trowel blazers or honorary men?. *J. Siam Soc.* 111(1): 139–158.
- HOANG, D. T., O. CHERNOMOR, A. VON HAESELER, B. Q. MINH, AND L. S. VINH. 2018. UFBoot2: Improving the Ultrafast Bootstrap Approximation. *Mol. Biol. Evol.* 35 (2): 518–522. <https://doi.org/10.1093/molbev/msx281>
- HORI, K., X. ZHOU, Y.H. YAN, Y. INOUE, AND N. MURAKAMI. 2018. Evidence for maternal ability in hybridization of apogamous fern species: *Dryopteris tsushimensis* K. Hori & N. Murak. and *D. subtsushimensis* K. Hori & N. Murak. (Dryopteridaceae), new tetraploid apogamous pteridophytes of hybrid origin from Tsushima, Japan. *Acta Phytotax. Geobot.* 69: 143–160. <https://doi.org/10.18942/apg.201809>
- HUIET, L., F.-W. LI, T.-T. KAO, J. PRADO, A. R. SMITH, E. SCHUETTPELZ, AND K. M. PRYER. 2018. A worldwide phylogeny of *Adiantum* (Pteridaceae) reveals remarkable convergent evolution in leaf blade architecture. *Taxon* 67 (3): 488–502.
- KALYAANAMORTHY, S., B. Q. MINH, T. K. F. WONG, A. VON HAESELER, AND L. S. JERMIN. 2017. ModelFinder: Fast model selection for accurate phylogenetic estimates. *Nature Methods* 14: 587–589. <https://doi.org/10.1038/nmeth.4285>
- KATO H. K., J. ROZEWICKI, AND K. D. YAMADA. 2019. MAFFT online service: multiple sequence alignment, interactive sequence choice and visualization. *Brief. Bioinform.* 20(4): 1160–1166. <https://doi.org/10.1093/bib/bbx108>

- KUO, L.-Y., T.-C. HSU, Y.-H. CHANG, Y. M. HUANG, AND W.-L. CHIOU. 2016. Systematics of the *Adiantum philippense* complex (Pteridaceae, Polypodiales) in Taiwan. *Phytotaxa* 263: 113–121. <https://doi.org/10.11646/phytotaxa.263.2.3>
- LARSSON, A. 2014. AliView: A fast and lightweight alignment viewer and editor for large datasets. *Bioinform.* 30(22): 3276–3278. <https://doi.org/10.1093/bioinformatics/btu531>
- LESTARI, W.S., B. ADJIE, T. JARUWATANAPHAN, Y. WATANO, AND M. PHARMAWATI. 2014. Molecular phylogeny of maidenhair fern genus *Adiantum* (Pteridaceae) from Lesser Sunda Islands Indonesia based on *rbcl* and *trnL-F*. *Reinwardtia* 14(1): 143–156. <https://doi.org/10.14203/reinwardtia.v14i1.409%20%20>
- LI, F.W., L.-Y. KUO, Y.-M. HUANG, W.-L. CHIOU, AND C. N. WANG. 2010. Tissue-direct PCR, a rapid and extraction-free method for barcoding of ferns. *Mol. Ecol. Resour.* 10: 92–95. <https://doi.org/10.1007/s00606-009-0188-2>
- LINDSAY, S. AND D. J. MIDDLETON. 2012–, continuously updated. *Ferns of Thailand, Laos and Cambodia*. Available from: <https://websites.rbge.org.uk/thaiferns/pages/lao-fern-list.htm> (accessed on 22 Oct. 2025)
- LU, J. M., J. WEN, S. LUTZ, Y. P. WANG, AND D. Z. LI. 2012. Phylogenetic relationships of Chinese *Adiantum* based on five plastid markers. *J. Pl. Res.* 125 (2): 237–249.
- MIYAZAKI, T., C. R. FRASER-JENKINS, D. T. VASQUES, P. SINAVONG, AND H. IKEDA. 2025. Five New Records of Pteridophytes from Laos. *J. Jap. Bot.* 100 (2): 149–153. <https://doi.org/10.51033/jjapbot.ID0230>
- MOES. 2024. *Education map of Laos 2020–2021*. (Thakhek district, Khammouan Province). [Lao language]. Available from: https://www.moes.edu.la/images/statistic/schoolmap/Maps_JPG2020_2021/12_khammouan/1201Thakhek.jpg
- NITTA, J. H., E. SCHUETTEPELZ, S. RAMÍREZ-BARAHONA, AND W. IWASAKI. 2022. An open and continuously updated fern tree of life. *Frontiers Pl. Sci. (Lausanne)* 13: 1–17. <https://doi.org/10.3389/fpls.2022.909768>
- NGUYEN, L.-T., H. A. SCHMIDT, A. VON HAESELER, AND B. Q. MINH. 2015. IQ-TREE: A fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Molec. Biol. Evol.* 32 (1):268–274. <https://doi.org/10.1093/molbev/msu300>
- PHONEPASEUTH, P., AND C.-W. LIN. 2025. *Begonia brunneosquamata* (sect. *Jackia*, Begoniaceae), A new species from limestone karsts of Bolikhamxai Province, Central Laos. *Phytotaxa* 720 (1): 67–74. <https://doi.org/10.11646/phytotaxa.720.1.6>
- PHONEPASEUTH, P., C. H. NGUYEN, D.H. VAN, P. SOULADETH, AND N. MURAKAMI. 2024. *Begonia* in Southern Laos with one new country and two new regional records. *Nat. Hist. Bull. Siam Soc.* 66 (1): 119–130.
- POWO. 2025–, continuously updated. *Plants of the World Online*. Available from: <https://powo.science.kew.org> (accessed on 20 Nov. 2025)
- QIAN, Y. Y. 1992. A new species of *Adiantum* from Yunnan. *Acta Bot. Austro Sin.* 8: 37–38.
- RAMBAUT, A. 2012. *FigTree, Version 1.4*. <http://tree.bio.ed.ac.uk/software/figtree>
- SOUVANNAKHOUMMANE, K., P. PHONEPASEUTH, P. SOULADETH, S. LANORSAVANH, S. TAGANE, AND N. TANAKA. 2025. Studies on the Gesneriaceae in Laos II: *Lysionotus calcicola*, a new species from limestone karsts in North and Central Laos, supported by morphological and molecular evidence. *Nord. J. Bot.* 2025: e04620. <https://doi.org/10.1002/njb.04620>
- SRIPENG, T., N. RONGDACH, P. NA-RANONG, T. JARUWATTANAPHAN, S. LANORSAVANH, C. LIMTAROPAS, AND J. SANGRATTANAPRASERT. 2024. *Adiantum membranifolium* (Pteridaceae), a new record of maidenhair fern from Laos. *Gard. Bull. Singapore* 76 (2): 277–283. [https://doi.org/10.26492/gbs76\(2\).2024-10](https://doi.org/10.26492/gbs76(2).2024-10)
- TABERLET, P., L. GIELLY, G. PAUTOU, AND J. BOUVET. 1991. Universal primers for amplification of three non-coding regions of chloroplast DNA. *Plant Mol. Biol.* 17: 1105–1109.
- TAGANE, S., P. SOULADETH, P. PHONEPASEUTH, T. VONGTHAVONE, D. KONGXAISAVATH, T. YAMAMOTO, AND N. MURAKAMI. 2025. *Tetrastigma fruticosum*, a new species of Vitaceae from limestone karst in central Laos. *Phytotaxa* 683 (1): 9–17. <https://doi.org/10.11646/phytotaxa.683.1.2>
- TAGANE, S., P. SOULADETH, D. KONGXAYSAVATH., S. RUEANGRUEA, S. SUDDÉE., Y. SUYAMA., E. SUZUKI, AND T. YAHARA. 2021. Two new species and 18 new records for the flora of Laos. *Thai For. Bull., Bot.* 49(1): 111–126.
- TAGAWA, M., AND K. IWATSUKI. 1985. Pteridophytes. in T. Smitinand, and K. Larsen (eds.), *Flora of Thailand*, Vol. 3, Part 2. Royal Forest Department, Bangkok.
- TARDIEU-BLOT, M., AND C. CHRISTENSEN. 1938. Ferns of Indo-Chine. *Notul. Syst. (Paris)* 6: 162–176.
- VONGTHAVONE, T., S. TAGANE, P. PHONEPASEUTH, K. SOUVANNAKHOUMMANE, AND P. SOULADETH. 2021. Twelve new records of pteridophytes from Bolaven Plateau, southern Laos. *Thai For. Bull., Bot.* 49 (2): 182–190. <https://doi.org/10.20531/tfb.2021.49.2.05>
- VONGTHAVONE, T., S. TAGANE, P. SOULADETH, K. SOUVANNAKHOUMMANE, P. PHONEPASEUTH, AND S. XAYYASITH. 2025. Two new species and five new distribution records of Pteridophytes from Phou Pha Marn Limestone Karst in Central Laos. *Kew Bull.* 80: 185–196. <https://doi.org/10.1007/s12225-024-10213-4>

- WANG, F.-H., J.-M. LU, J. WEN, A. EBIHARA, AND D. Z. LI. 2016. Applying DNA barcodes to identify closely related Species of ferns: A case study of the Chinese *Adiantum* (Pteridaceae). *PLoS ONE* 11(9): e0160611. <https://doi.org/10.1371/journal.pone.0160611>
- YONEOKA, K., T. FUJIWARA, T. KATAOKA, K. HORI, A. EBIHARA, AND N. MURAKAMI. 2024. Morphological and functional evolution of gametophytes in epilithic *Hymenasplenium murakami-hatanakae* (Aspleniaceae): The fifth family capable of producing the independent gametophytes. *J. Plant Res.* 137: 815–828. <https://doi.org/10.1007/s10265-024-01553-0>
- ZHANG, G. M., W. B. LIAO, M. Y. DING, Y. X. LIN, Z. H. WU, X. C. ZHANG, S. Y. DONG, J. PRADO, M. G. GILBERT, G. YATSKIEVYCH, T. A. RANKER, E. A. HOOPER, E. R. ALVERSON, J. S. METZGAR, A. M. FUNSTON, S. MASUYAMA, AND M. KATO. 2013. Pteridaceae. Pages 169–256 in Z. Y. Wu, P. H. Raven, and D. Y. Hong (eds), *Flora of China* Vols 2–3 (Pteridophytes). Science Press, Beijing & Missouri Botanical Garden Press, St. Louis.
- ZHANG, L., N. T. LU, X.-M. ZHOU, Z. ZHOU, K. THEPKAYSON, T. T. LUONG, AND L.-B. ZHANG. 2024. Exploring the diversity of the Java fern genus *Leptochilus* (Polypodiaceae) in the Indo-Burma biodiversity hotspot. *Phytotaxa* 659 (3): 213–235. <https://doi.org/10.11646/phytotaxa.659.3.1>

Appendix 1. Pteridophyte taxa recently documented from Laos (2021–2025).

Family	Taxon	Region	Province	Life form and habitat	Distribution	References
ATHYRIACEAE	<i>Comopteris opaca</i> (D.Don) Tagawa	South	Champasak	Terrestrial on wet ground in hill evergreen forest	India, Bhutan, Nepal, China, Japan, Thailand, Laos, Vietnam, Indonesia	TAGANE <i>ET AL.</i> (2021)
	<i>Diplazium platyklamys</i> C.Chr.	South	Champasak	Terrestrial in hill evergreen forest	Laos and Vietnam	TAGANE <i>ET AL.</i> (2021)
	<i>Asplenium perakense</i> C.G.Matthew & Christ	South	Champasak	Lithophytic on semi-shaded mossy moist rocks in wet evergreen forest	Thailand, Laos, and Malaysia	VONGTHAVONE <i>ET AL.</i> (2021)
	<i>Asplenium simonstanum</i> Hook.	South	Champasak	Epiphytic on moss-covered tree trunks in wet evergreen forest	India, China, Thailand, and Laos	VONGTHAVONE <i>ET AL.</i> (2021)
	<i>Asplenium tenerum</i> G.Forst.	South	Champasak	Lithophytic on semi-shaded rocks and Epiphyte on tree trunks in dense montane conifer forest	India, Sri Lanka, Japan (Ogasawara), Korea, Laos, Myanmar, Vietnam, Cambodia, Malaysia, Indonesia, Philippines, Pacific islands	TAGANE <i>ET AL.</i> (2021)
HYMENASPLENIACEAE	<i>Hymenasplenium cheilosorum</i> (Kunze ex Mett.) Tagawa	South	Champasak	Terrestrial on wet ground in hill evergreen forest	India, Bhutan, Nepal, Sri Lanka, China, Japan, Myanmar, Thailand, Laos, Vietnam, Malaysia, Indonesia, Philippines	TAGANE <i>ET AL.</i> (2021)

Appendix 1 (continued).

Family	Taxon	Region	Province	Life form and habitat	Distribution	References
DENNSTAEDTIACEAE	<i>Histiopteris incisa</i> (Thunb.) J.Sm.	South	Champasak	Terrestrial on exposed and humus-rich slopes in wet evergreen forest	Pantropical and Laos	VONGTHAVONE ET AL. (2021)
	<i>Microlepia calvescens</i> (Hook.) C.Presl	North	Oudomxay	Terrestrial on reddish soil site on road side, forest margins, secondary forest	India, E. Himalaya, S. China, Upper Myanmar, Thailand, Laos, Vietnam, Indonesia, and Taiwan	MIYAZAKI ET AL. (2025)
DRYOPTERIDACEAE	<i>Arachniodes pseudoassamica</i> Ching	South	Champasak	Terrestrial in semi-shaded under-story in wet evergreen forest	China, and Laos	VONGTHAVONE ET AL. (2021)
	<i>Polystichum attenuatum</i> Tagawa & K.Iwats.	South	Champasak	Terrestrial on humus-rich ground in wet evergreen forest	India, Myanmar, China, Taiwan, Thailand, Laos, and Vietnam	VONGTHAVONE ET AL. (2021)
HYMENOPHYLLACEAE	<i>Crepidomanes christii</i> (Copel.) Copel.	South	Champasak	Epiphytic on tree trunks in lowland evergreen forest	India, Thailand, Laos, Malaysia, and Indonesia, and Philippines	VONGTHAVONE ET AL. (2021)
HYPODEMATIACEAE	<i>Leucostegia truncata</i> (D.Don) Fraser-Jenk.	South	Champasak	Epiphytic on moss-covered bed-rock in wet evergreen forest	India, Bhutan, Nepal, Sri Lanka, China, Taiwan, Myanmar, Thailand, Laos, Vietnam, Cambodia, Malaysia, Indonesia, Philippines, and New Guinea	VONGTHAVONE ET AL. (2021)

Appendix 1 (continued).

Family	Taxon	Region	Province	Life form and habitat	Distribution	References
LINDSAEACEAE	<i>Lindsaea orbiculata</i> (Lam.) Mett. ex Kuhn	North	Oudomxay	Terrestrial on reddish soil site on road cutting, half shade	NE India, S China, Myanmar Laos, Thailand, Cambodia, W & C Malasia, Taiwan, S Japan	MIYAZAKI <i>ET AL.</i> (2025)
LYCOPODIACEAE	<i>Palhinhaea crassifolia</i> (Spring) Fraser-Jenk. & Kholia	North	Oudomxay	Terrestrial on reddish soil site on sunny roadside	Laos, India, SE Asia	MIYAZAKI <i>ET AL.</i> (2025)
PLAGIOGYRIACEAE	<i>Plagiogyria adnata</i> (Blume) Bedd.	South	Champasak	Terrestrial in primary dense montane conifer forest, near streams	India, China, Japan, Myanmar, Thailand, Laos, Vietnam, Malaysia (Borneo, Peninsula), Indonesia (Java, Sumatra), Philippines (Luzon)	TAGANE <i>ET AL.</i> (2021)
POLYPODIACEAE	<i>Drynaria deusavanhii</i> Vongthavone & Tagane	Central	Kham- mouane	Lithophytic on limestone karst/ Epiphyte on tree trunk.	Endemic to Laos	VONGTHAVONE <i>ET AL.</i> (2025)
	<i>Grammitis barathrophylla</i> (Baker) Christenh.	South	Champasak	Epiphytic on lower part of mossy tree trunks in wet evergreen forest	China, Thailand, Laos, Vietnam, Cambodia, Malaysia, Indonesia, Philippines, and New Guinea.	VONGTHAVONE <i>ET AL.</i> (2021)
	<i>Lepisorus henryi</i> (Hieron. ex C.Chr.) Li Wang	South	Champasak	Epiphytic on tree trunks in wet evergreen forests	India, Bhutan, Nepal, China, Thailand, Laos, and Vietnam	VONGTHAVONE <i>ET AL.</i> (2021)

Appendix 1 (continued).

Family	Taxon	Region	Province	Life form and habitat	Distribution	References
Fungi	<i>Lepisorus spicatus</i> (L.f.) Li Wang	South	Champasak	Epiphytic on tree trunks in wet evergreen forests	Tropical Asia and Laos.	VONGTHAVONE ET AL. (2021)
	<i>Leptochilus bolikhamsatensis</i> Liang Zhang, Thepkaysone & Z.Zhou	Central	Bo-likhamxai	Lithophytic on limestone rocks in disturbed rainforest	Endemic to Laos	ZHANG ET AL. (2024)
	<i>Leptochilus hemionitideus</i> (Wall. ex C.Presl) Noot.	South	Champasak	Terrestrial on wet ground in hill evergreen forest	India, Bhutan, Nepal, China, Japan, Thailand, and Laos	TAGANE ET AL. (2021)
	<i>Leptochilus khammouanensis</i> Liang Zhang, Thepkaysone & Z.Zhou	Central	Kham-mouane	Lithophytic on limestone rock in disturbed primary forests	Endemic to Laos	ZHANG ET AL. (2024)
	<i>Leptochilus luangprabangensis</i> Liang Zhang, Thepkaysone & Z.Zhou	North	Luang Prabang	Hemiepiphytic on a small tree trunk, the roots extended into the soil below, in limestone mountain	Endemic to Laos	ZHANG ET AL. (2024)
	<i>Leptochilus wusugongii</i> Liang Zhang & Li Bing Zhang	North	Luang Prabang	Lithophytic on limestone in the primary forest	Endemic to Laos	ZHANG ET AL. (2024)
	<i>Microsorium steerei</i> (Harr.) Ching	Central	Kham-mouane	Epiphytic on tree trunk in limestone karst forest	China, Laos, Vietnam, and Taiwan.	VONGTHAVONE ET AL. (2025)
	<i>Pyrrosia flocculosa</i> (D.Don) Ching	Central	Kham-mouane	Epiphytic on tree trunk in limestone karst forest	India, Nepal, Bhutan, Bangladesh, China, Myanmar, Thailand, Laos, and Vietnam	VONGTHAVONE ET AL. (2025)
	<i>Selliguea oxyloba</i> (Wall. ex Kunze) Fraser-Jenk.	South	Champasak	Lithophytic on exposed bedrocks beside open grassland or epiphytic on moss-covered tree trunks	India, Nepal, Myanmar, China, Thailand, Laos, and Vietnam	VONGTHAVONE ET AL. (2021)

Appendix 1 (continued).

Family	Taxon	Region	Province	Life form and habitat	Distribution	References
PTERIDACEAE	<i>Tricholepidium normale</i> (D. Don) Ching	South	Champasak	Epiphytic on mossy tree trunk in wet evergreen forest	India, Bhutan, Nepal, Myanmar, China, Japan, Thailand, Laos, Vietnam, Malaysia, and Indonesia.	VONGTHAVONE ET AL. (2021)
	<i>Adiantum membranifolium</i> S.Linds. & Suksathan	Central	Khammouane	Lithophytic on limestone karst/wall	Laos and Thailand	SRIPENG ET AL. (2024)
	<i>Adiantum menglianense</i> Y.Y.Qian	North	Luang Prabang	Terrestrial in limestone karst forest	China, Laos, and Taiwan	This study
	<i>Antrophyum formosanum</i> Hieron.	Central	Khammouane	Epiphytic on lower tree trunk, in wet evergreen forests	Laos, Taiwan, and Japan	VONGTHAVONE ET AL. (2025)
	<i>Hemionitis fragilis</i> (Hook.) Christenh.	Central	Khammouane	Lithophytic on limestone karst	Africa, Europe, and Asia (China, Myanmar, Thailand, Laos and Vietnam)	VONGTHAVONE ET AL. (2025)
	<i>Hemionitis khammouanensis</i> Vongthavone & Phonep.	Central	Khammouane	Lithophytic on limestone karst	Endemic to Laos	VONGTHAVONE ET AL. (2025)
	<i>Pteris taiwanensis</i> Ching	North	Oudomxay	Terrestrial on the side of stream, edge of forest	Assam, Nepal, East Himalaya, Laos, and Taiwan	MIYAZAKI ET AL. (2025)
	<i>Amblovenatum decorum</i> (Domin) S.Linds. & D.J.Middleton	Central	Khammouane	Terrestrial near Waterfall	Africa, Asia (Sri Lanka, Thailand, Laos, Vietnam, Cambodia, Malesia) and Australia	VONGTHAVONE ET AL. (2025)
	<i>Metahelipteris laxa</i> (Franch. & Sav.) Ching	North	Oudomxay	Terrestrial in forest margin in secondary forest	China, Thailand, Laos, Japan, S. Korea, and Taiwan	MIYAZAKI ET AL. (2025)
THELYPTERIDACEAE						